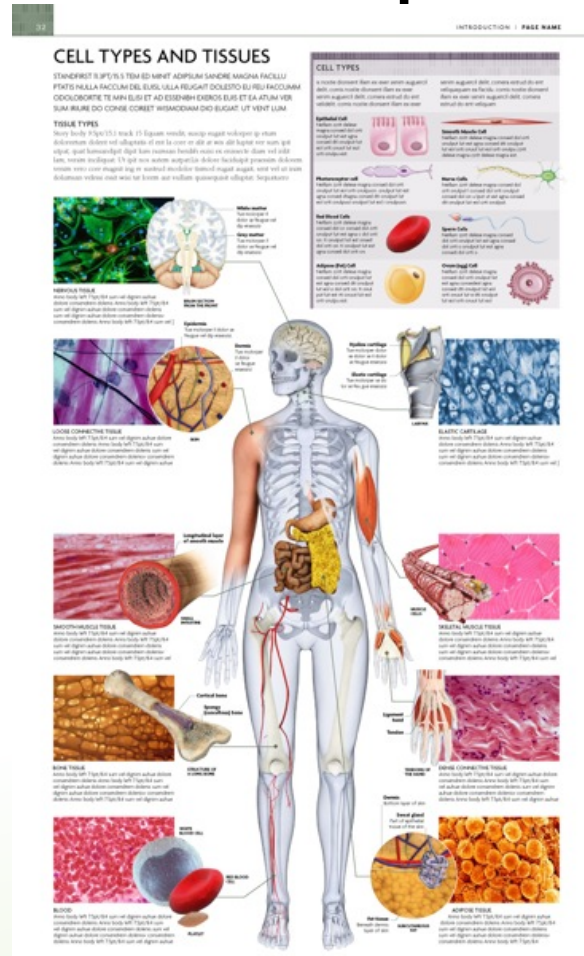


RNA-seq Introduction

Promises and pitfalls

Enabler for Life Sciences

RNA gives information on which genes that are expressed



How DNA get transcribed to RNA (and sometimes then translated to proteins) varies between e. g.

- Tissues
- Cell types
- Cell states
- Individuals
- Cells

RNA gives information on which genes that are expressed

How DNA get transcribed to RNA (and sometimes then translated to proteins) varies between e. g.



-Tissues

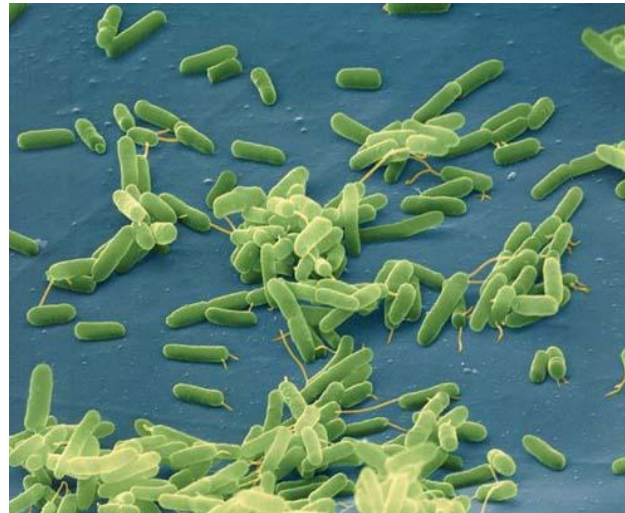
-Cell types

-Cell states

-Individuals

RNA gives information on which genes that are expressed

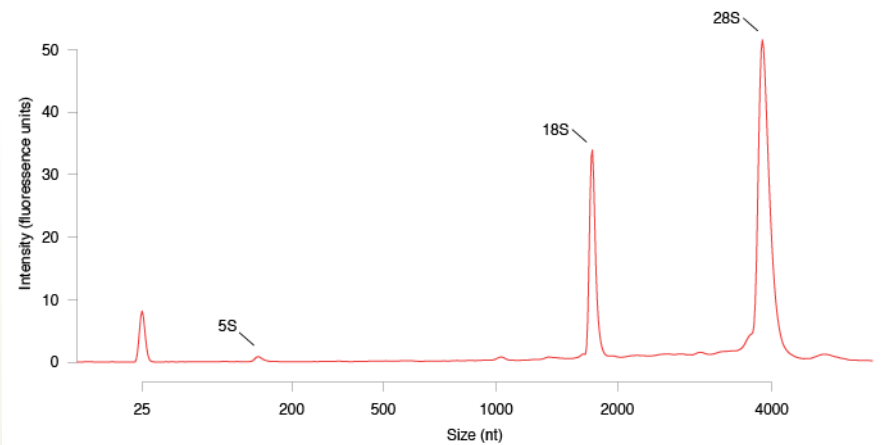
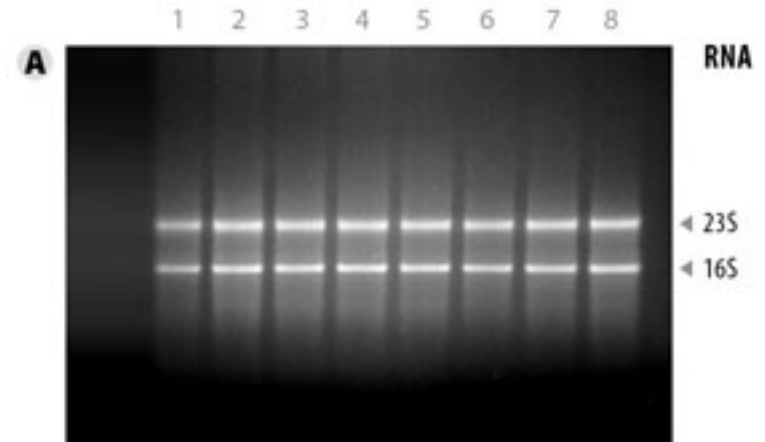
How DNA get transcribed to RNA (and sometimes then translated to proteins) varies between e. g.



- Tissues
- Cell types
- Cell states
- Individuals


RNA flavors (pre sequencing era)

- House keeping RNAs
 - rRNAs, tRNAs, snoRNAs, snRNAs, SRP RNAs, catalytic RNAs (RNase E)
- Protein coding RNAs
 - (1 coding gene ~ 1 mRNA)
- Regulatory RNAs
 - Few rare examples



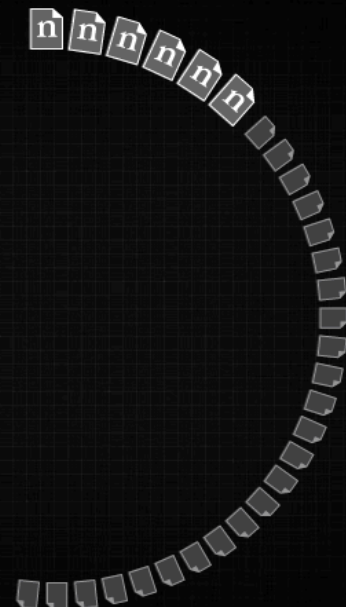
nature
ENCODE explorer

THREADS



PAPERS

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ENCODE, the Encyclopedia of DNA Elements, is a project funded by the National Human Genome Research Institute to identify all regions of transcription, transcription factor association, chromatin structure and histone modification in the human genome sequence.

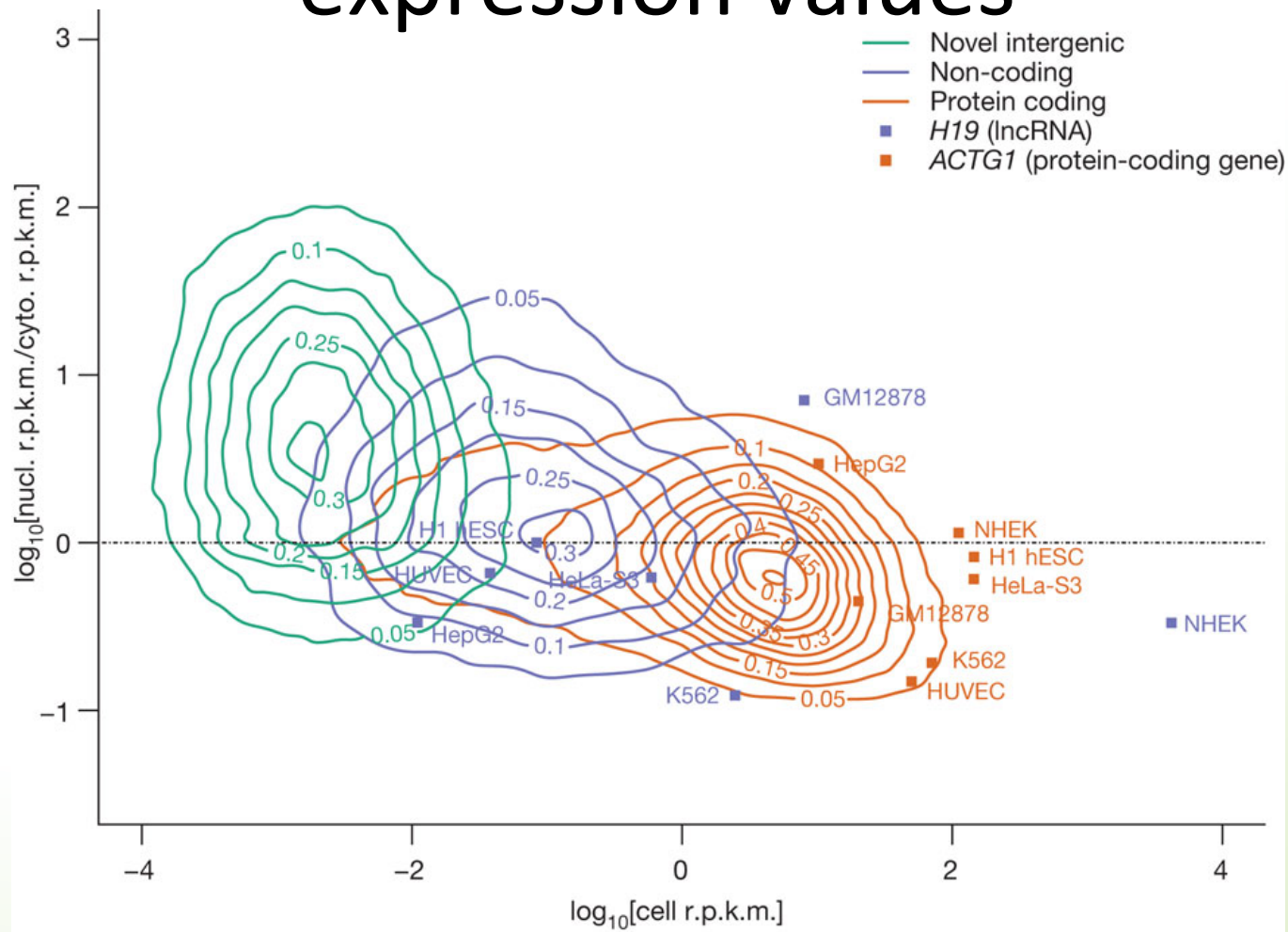
ENCyclopedia Of Dna Elements

ENCODE By the Numbers

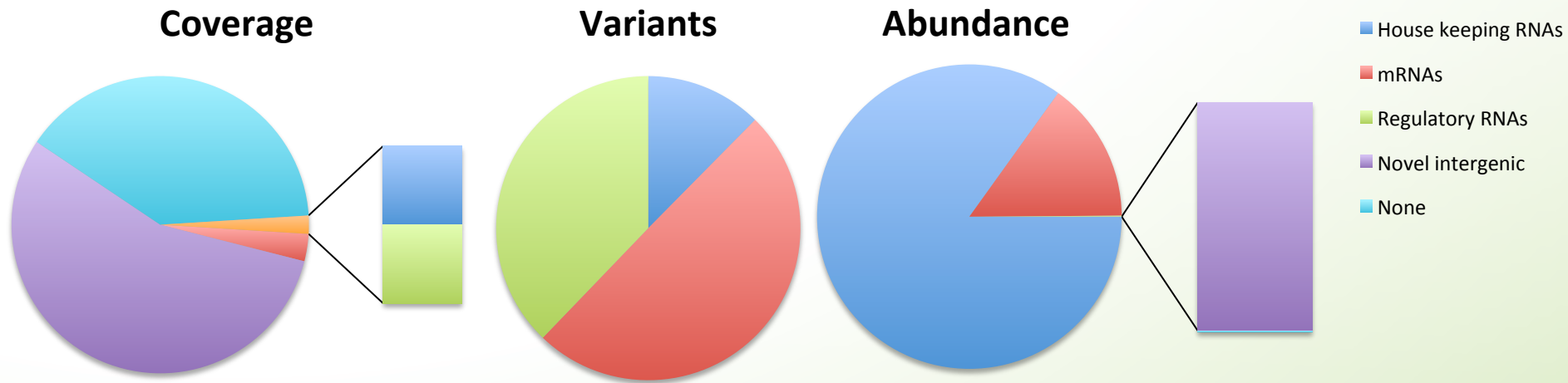
- 147** cell types studied
- 80%** functional portion of human genome
- 20,687** protein-coding genes
- 18,400** RNA genes
- 1640** data sets
- 30** papers published this week
- 442** researchers
- \$288 million** funding for pilot technology, model organism, and current

Cumulatively, we observed a total of 62.1% and 74.7% of the human genome to be covered by either processed or primary transcripts, respectively, with no cell line showing more than 56.7% of the union of the expressed transcriptomes across all cell lines.

Different kind of RNAs have different expression values



What defines RNA depends on how you look at it



Defining functional DNA elements in the human genome

- Statement
 - A priori, we should not expect the transcriptome to consist exclusively of functional RNAs.
- Why is that
 - Zero tolerance for errant transcripts would come at high cost in the proofreading machinery needed to perfectly gate RNA polymerase and splicing activities, or to instantly eliminate spurious transcripts.
 - In general, sequences encoding RNAs transcribed by noisy transcriptional machinery are expected to be less constrained, which is consistent with data shown here for very low abundance RNA
- Consequence
 - Thus, one should have high confidence that the subset of the genome with large signals for RNA or chromatin signatures coupled with strong conservation is functional and will be supported by appropriate genetic tests.
 - In contrast, the larger proportion of genome with reproducible but low biochemical signal strength and less evolutionary conservation is challenging to parse between specific functions and biological noise.

This is of course not without an debate

OPEN ACCESS Freely available online PLOS BIOLOGY

Most "Dark Matter" Transcripts Are Associated With

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Perspective

The Reality of Pervasive Transcription

Michael Rinn

OPEN ACCESS Freely available online PLOS BIOLOGY

Perspective

Comments on van Bakel et al. (2011) Response to "The Reality of Pervasive Transcription"

Comments by Mike Clark

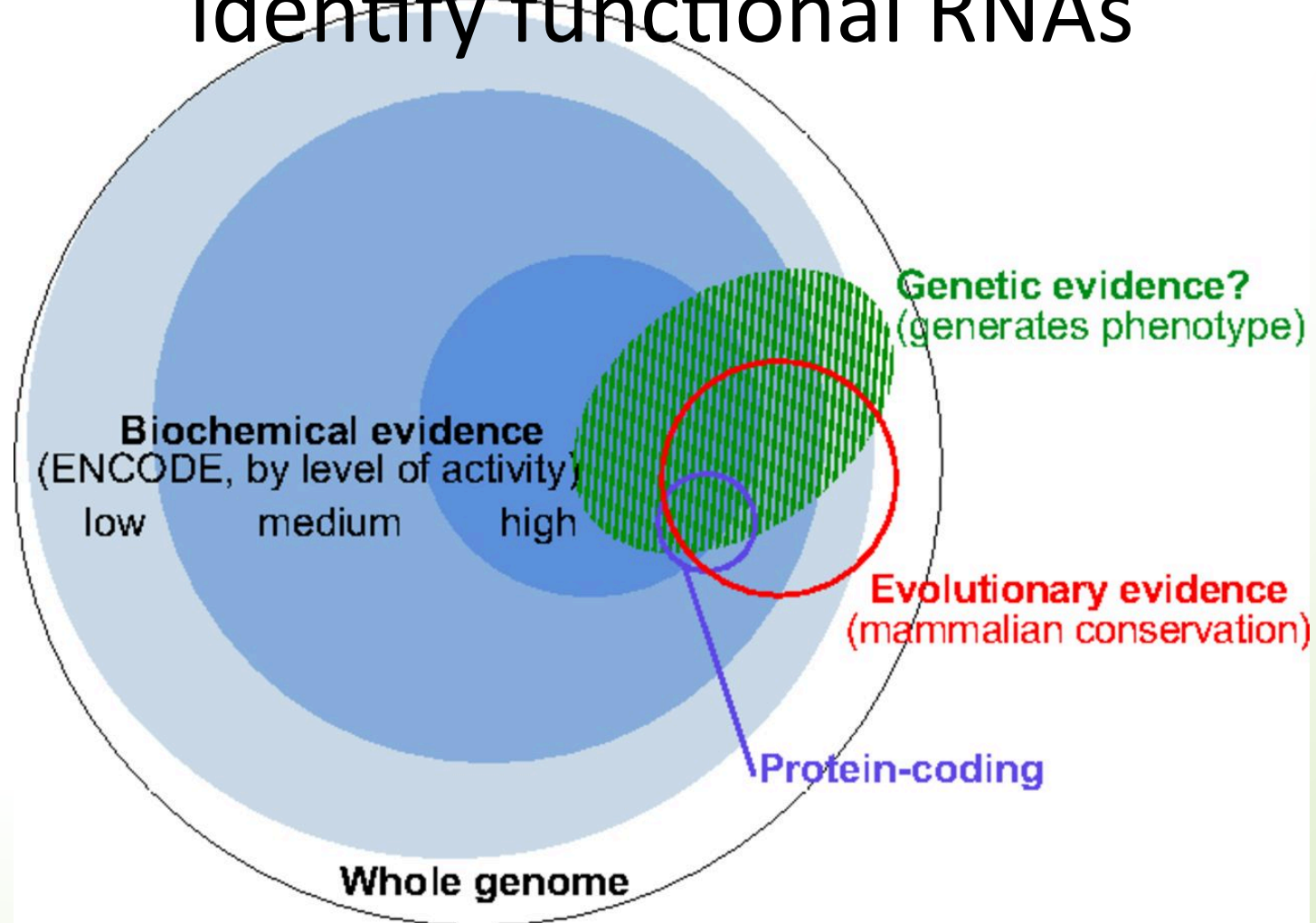
Van Bakel et al. 2011 (vB 11) have published their reply to our critique of their paper van Bakel et al. 2010 (vB 10).

Firstly lets briefly review some of our main criticisms of vB 10:

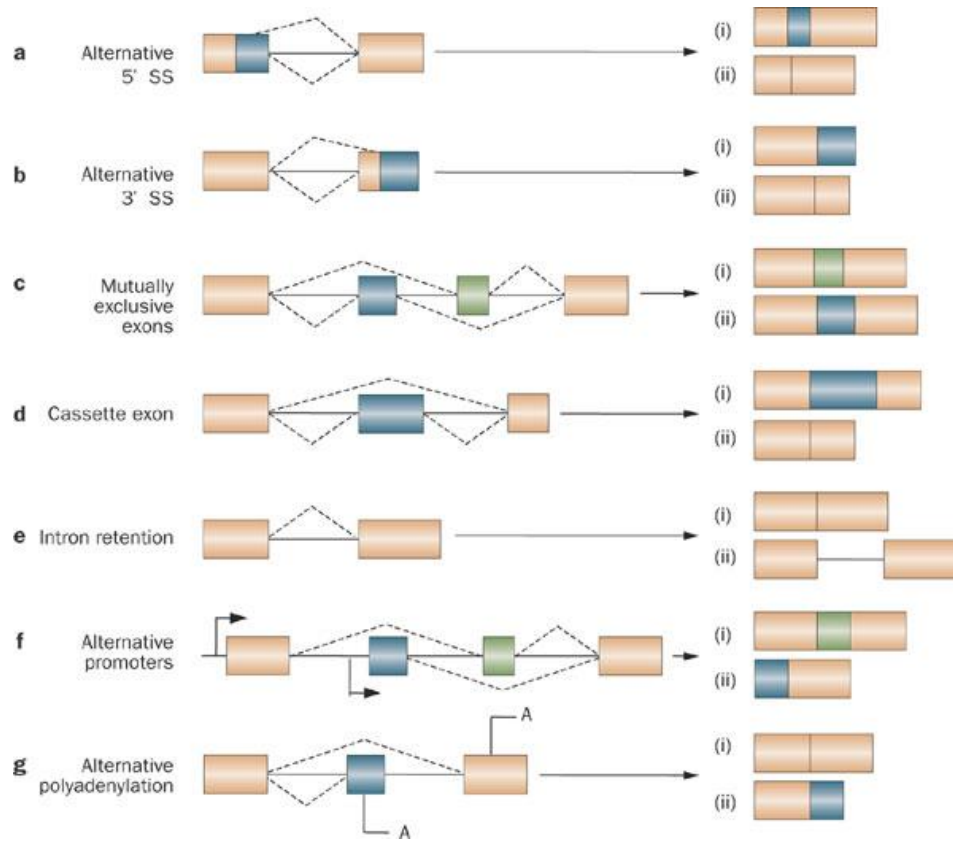
1. vB 10 didn't properly consider previous evidence for pervasive transcription (especially that from cDNA analysis in the mouse) when claiming the genome was not as transcribed as previous evidence was unreliable due to false positives.
2. vB 10 incorrectly conflated pervasive transcription with the relative abundance of transcripts when the correct (and known) definition was the amount of the genome that was transcribed.
3. The tiling arrays vB 10 performed and then used to claim that previous array studies suffered from high false positives were atypical and lacked any validation of the false positive rate.
4. The RNA sequencing carried out by vB 10 was severely limited in its ability to address the question of pervasive transcription. The depth of sequencing was too shallow to detect rare transcripts and the assembly of what was found into transcripts was poor. Since it couldn't detect and/or characterize rare transcripts this meant it couldn't even identify the full set of transcripts.
5. vB 10 claimed that low level intergenic transcription may be due to "random initiation events" and/or transcriptional "byproducts" (ie: transcription noise), when the limitation was to properly differentiate between this and genuine transcripts under their detection threshold.

Legend:
Novel intergenic (green)
Coding (blue)
RNA (protein-coding gene) (red)

Biochemical evidence not enough to identify functional RNAs



One gene many different mRNAs

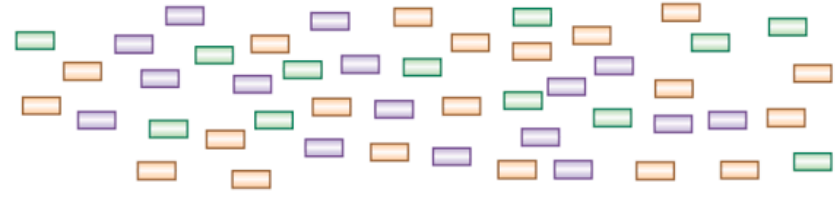


- RNA seq course

The RNA seq course

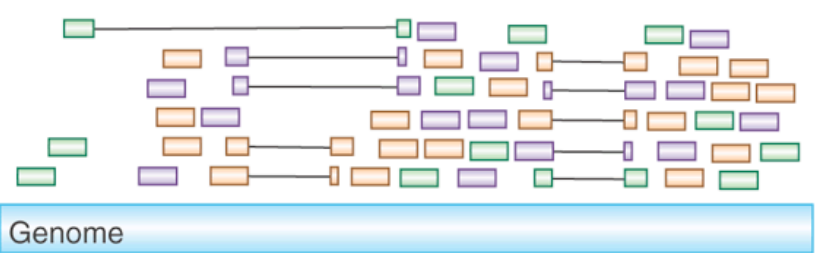
- From RNA seq to reads
- Mapping reads programs
- Transcriptome reconstruction using reference
- Transcriptome reconstruction without reference
- QC analysis
- sRNA analysis
- Differential expression analysis
 - mRNAs
 - miRNAs
- Genome annotation using RNA and other sources
- Differential expression using multi variate analysis
- RNA long read analysis

From RNA to short reads

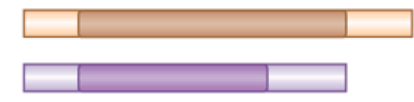
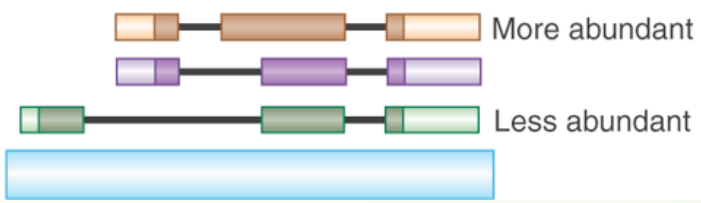


Align reads to genome

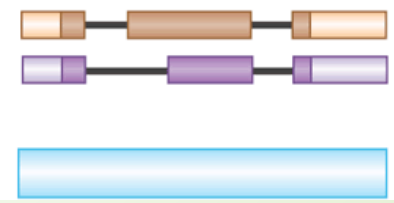
Assemble transcripts *de novo*



Assemble transcripts from spliced alignments



Align transcripts to genome



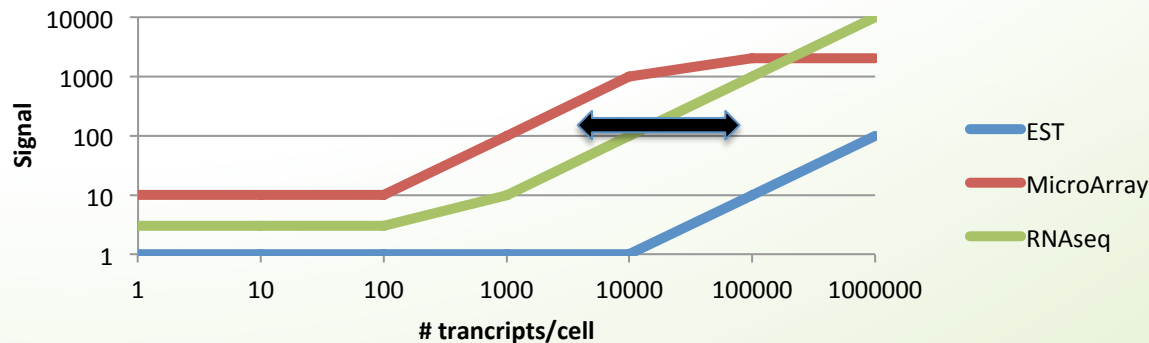
Promises and pitfalls

Long reads

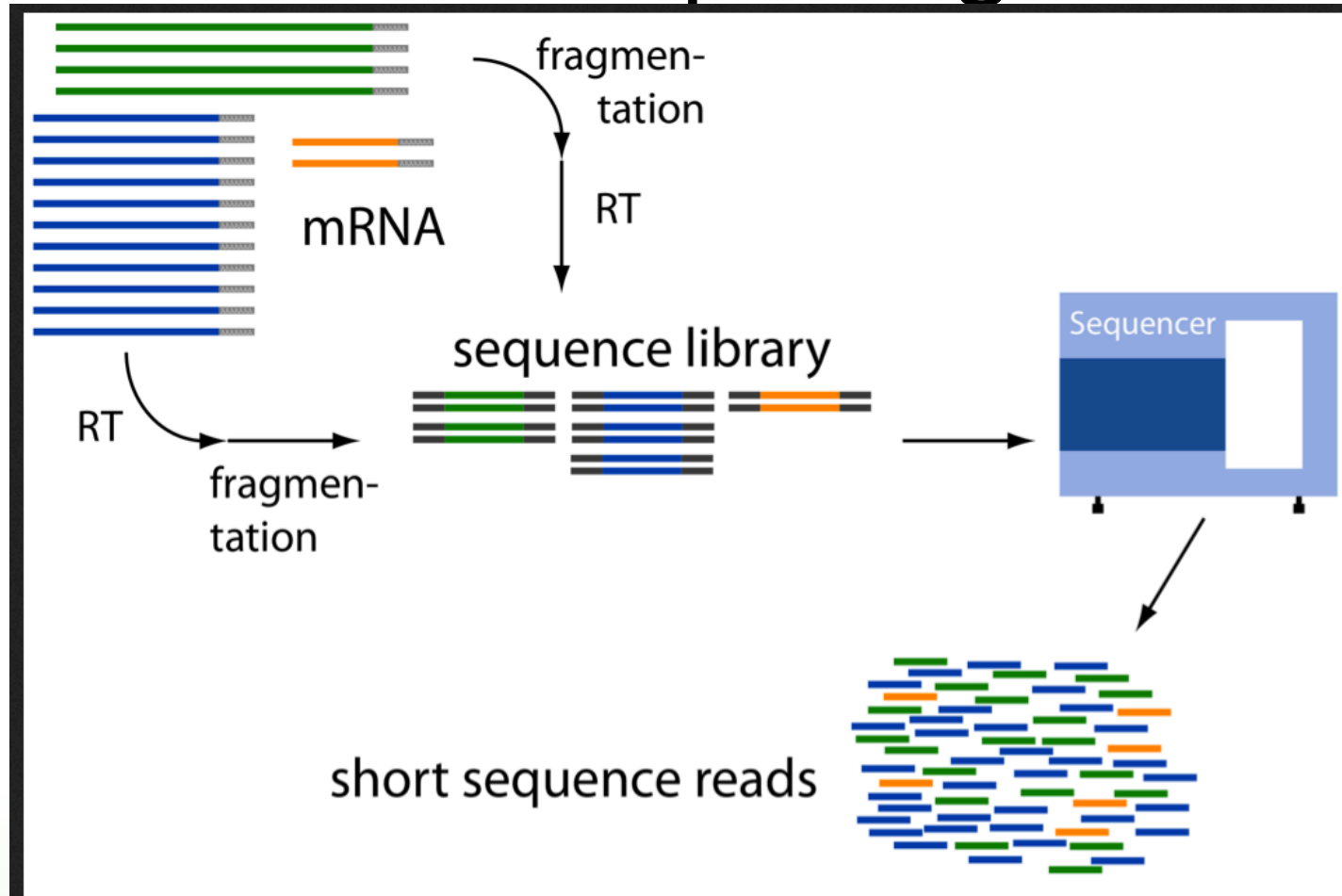
Micro Arrays

RNAseq

- | | | |
|------------------------------------|---|--------------------------------|
| • Low throughput (-) | • High throughput (+) | • High throughput (+) |
| • Complete transcripts (+) | • Only known sequences (-) | • Fractions of transcripts (-) |
| • Only highly expressed genes (--) | • Limited dynamic range (-) | • Full dynamic range (+/-) |
| • Expensive (-) | • Cheap (+) | • Unlimited dynamic range (+) |
| • Low background noise (+) | • High background noise (-) | • Cheap (+) |
| • Easy downstream analysis (+) | • Not strand specific (-) | • Low background noise (+) |
| | • Well established downstream methods (+) | • Strand specificity (+) |
| | | • Re-sequencing (+) |

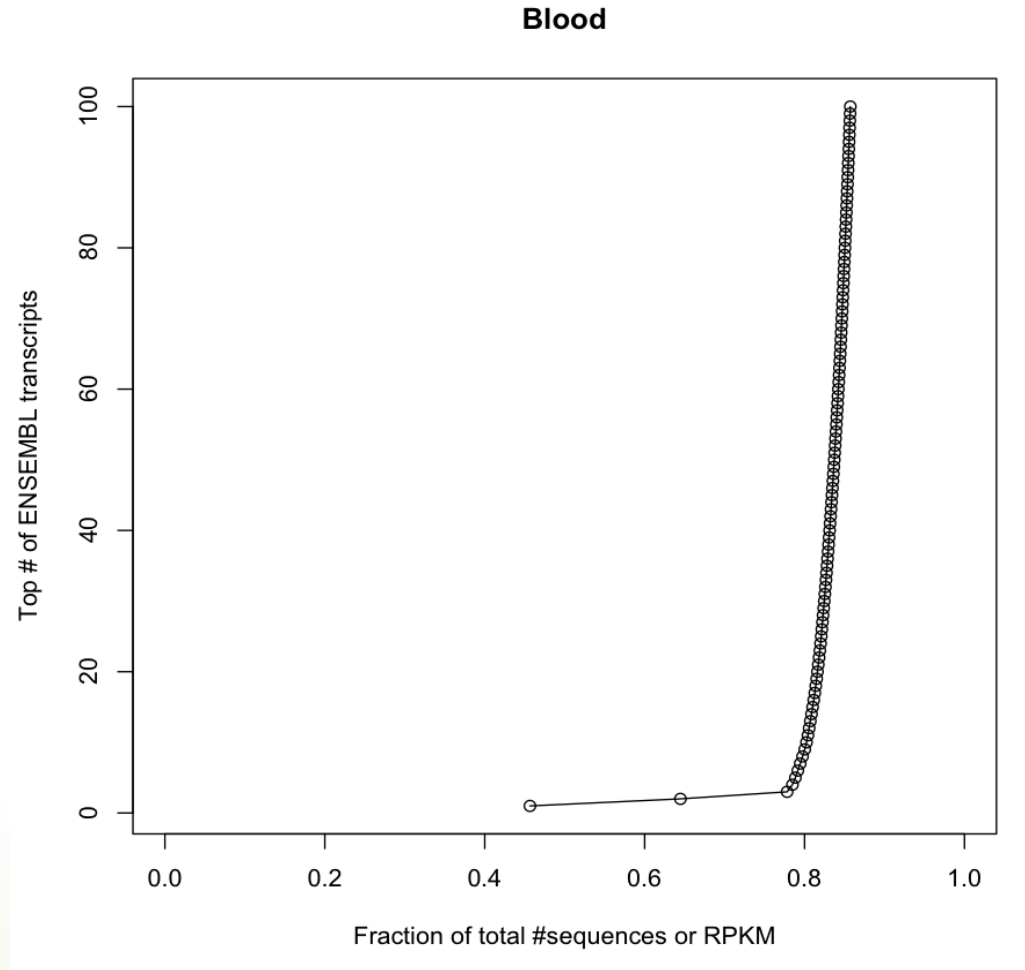


How are RNA-seq data generated?



Sampling process

RNA seq reads correspond directly to abundance of RNAs in the sample



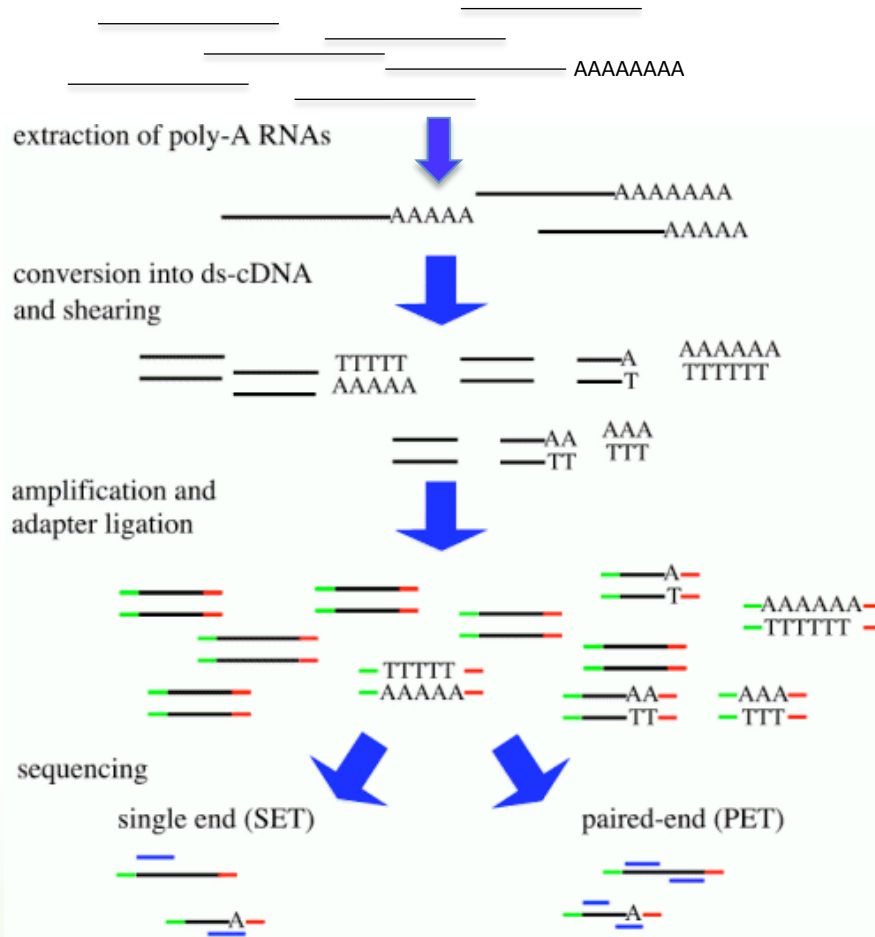
RNA to reads

RNA->

enrichments ->

library ->

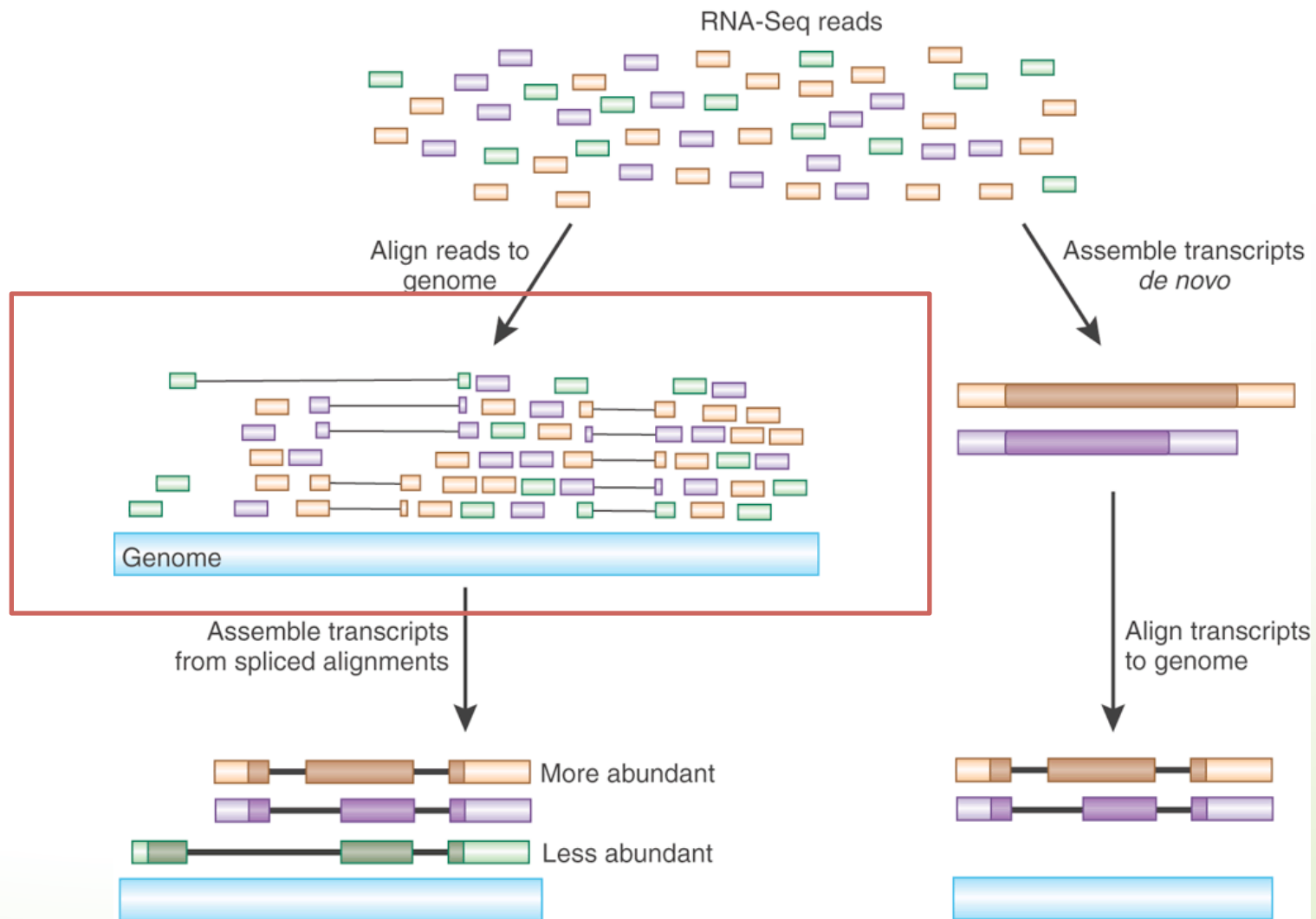
reads ->



PolyA (mRNA)
 RiboMinus (- rRNA)
 Size <50 nt (miRNA)

Size of fragment
 Strand specific
 5' end specific
 3' end specific

Single end (1 read per fragment)
 Paired end (2 reads per fragment)

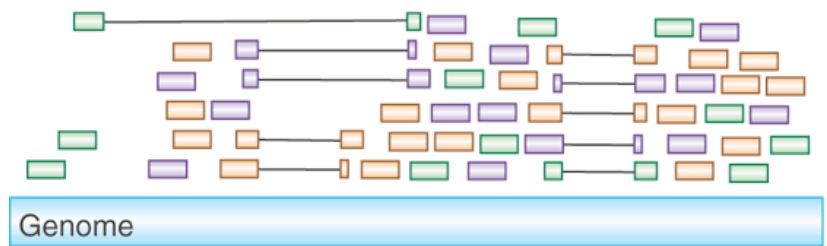




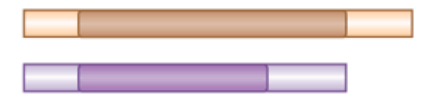
RNA-Seq reads

Align reads to genome

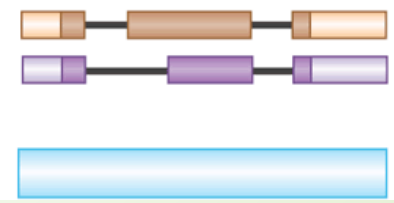
Assemble transcripts *de novo*



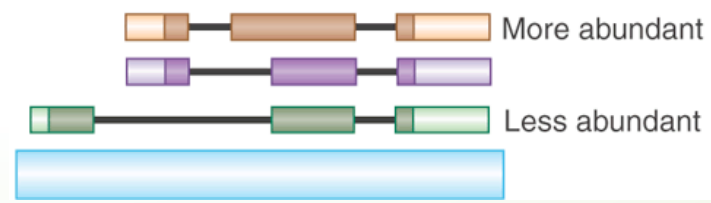
Assemble transcripts from spliced alignments

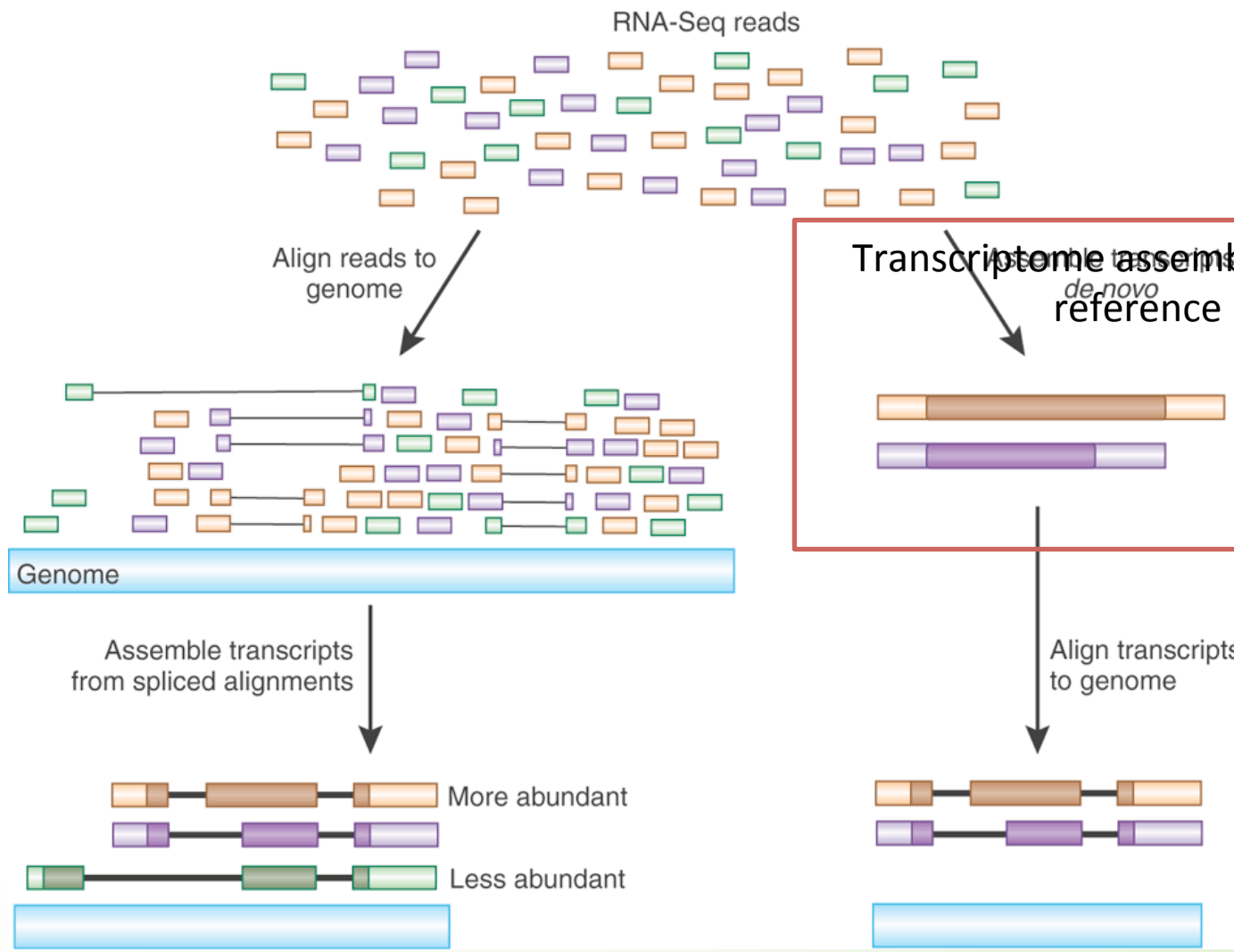


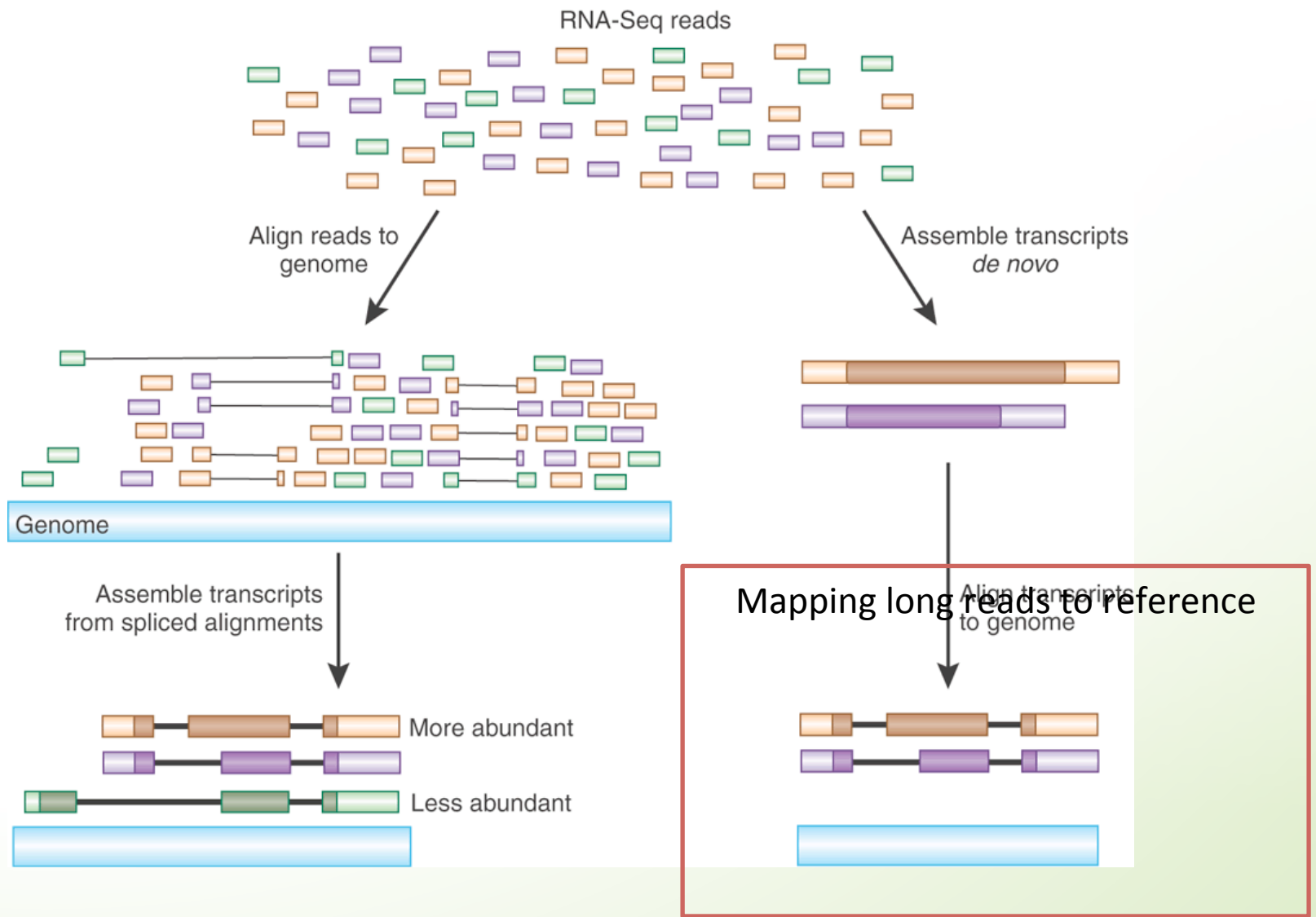
Align transcripts to genome



Transcriptome assembly using reference







Quality control

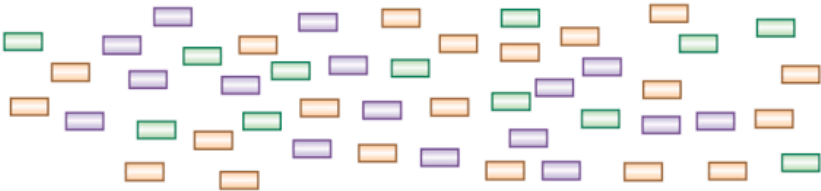
-samples might not be what you think they are

- Experiments go wrong
 - 30 samples with 5 steps from samples to reads has 150 potential steps for errors
 - Error rate 1/100 with 5 steps suggest that one of every 20 samples the reads does not represent the sample
- Mixing samples
 - 30 samples with 5 steps from samples to reads has ~24M potential mix ups of samples
 - Error rate 1/ 100 with 5 steps suggest that one of every 20 sample is mislabeled
- Combine the two steps and approximately one of every 10 samples are wrong

RNA QC

RNA-Seq reads

Read quality



Align reads to genome

Assemble transcripts *de novo*

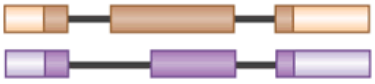
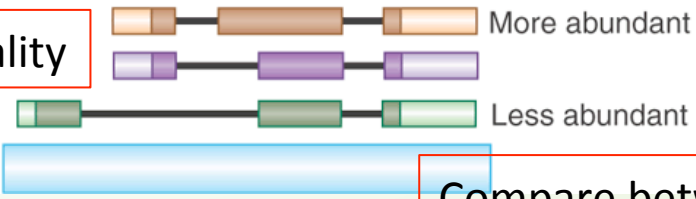
Mapping statistics



Assemble transcripts from spliced alignments

Align transcripts to genome

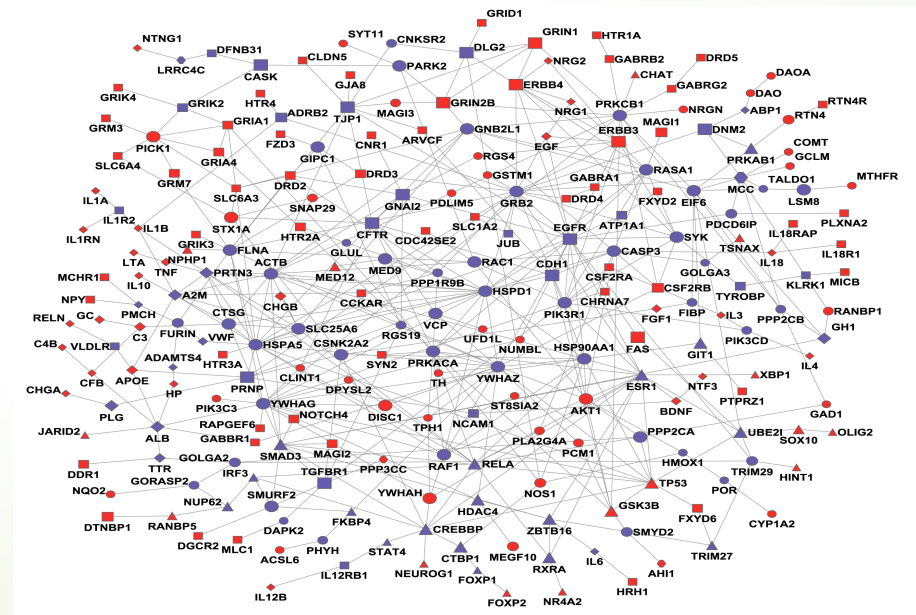
Transcript quality



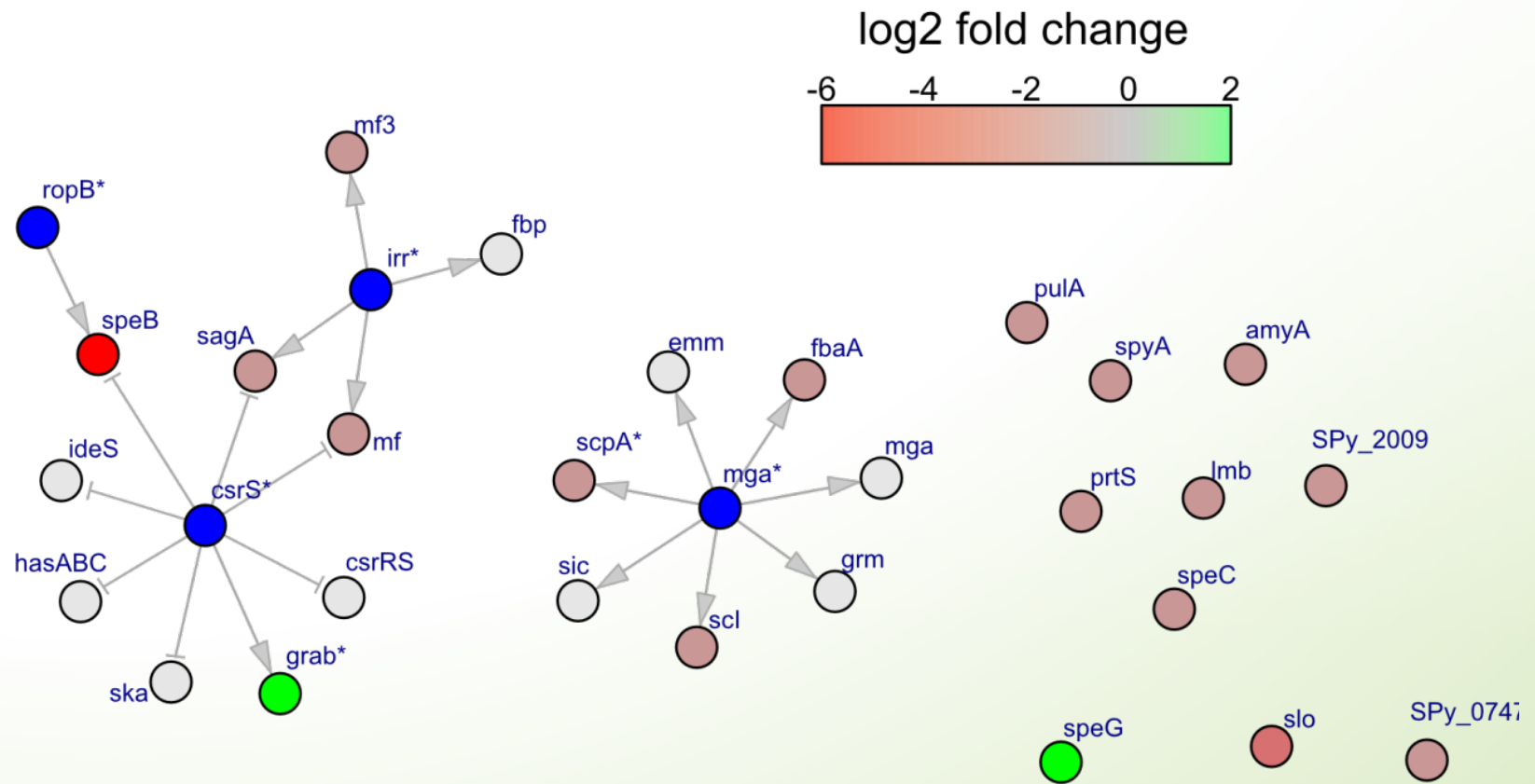
Compare between samples

Differential expression analysis using univariate analysis

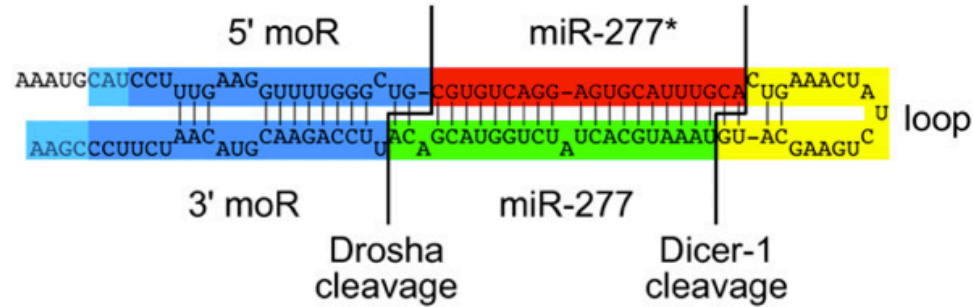
Typically **univariate** analysis (one gene at a time) – even though we know that genes are not independent



Gene set analysis and data integration



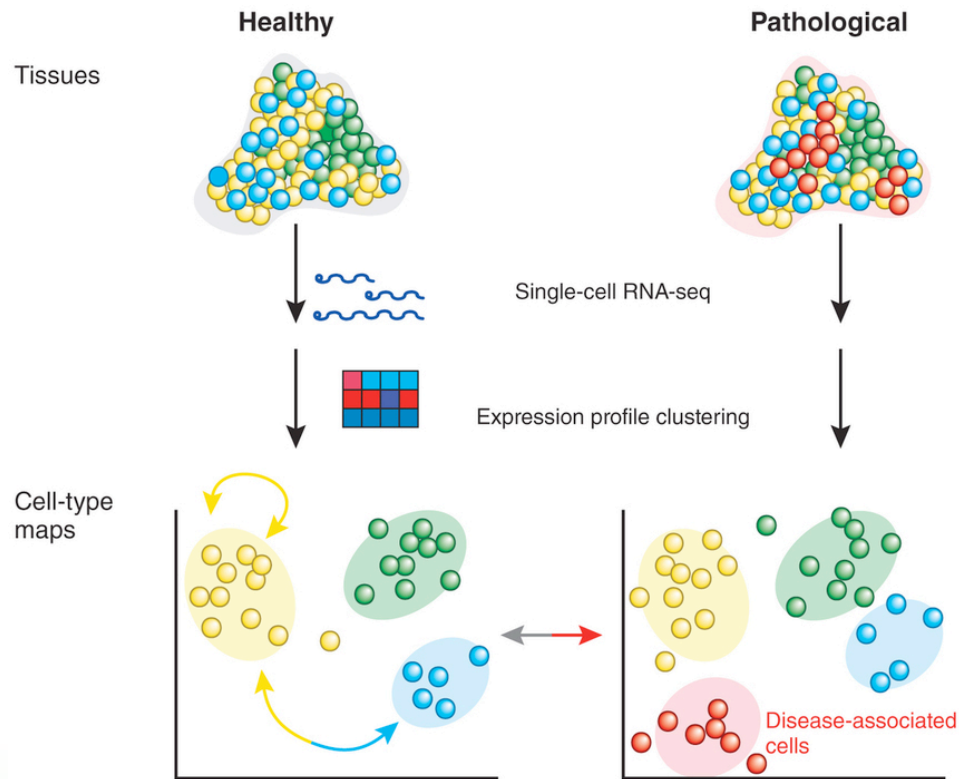
microRNA analysis (Jakub)



5' moR	miR-277*	loop	miR-277	3' moR	len	reads
AAATGCATCCTTTGAAGGTTTTGGGCTG	CGTGTCAGGAGTGCATTTGCACTG	GAAACTATCTGAAGCATG	TAAATGCACTATCTGGTACGACAT	TCCAGAACGTACAATCTTCCCGAA	23	1016281
-----	-----	-----	TAAATGCACTATCTGGTACGACA	-----	22	327660
5' fixed	-----	-----	TAAATGCACTATCTGGTACGAC	-----	21	217490
-----	CGTGTCAGGAGTGCATTTGCA	5' fixed	TAAATGCACTATCTGGTACGA	-----	21	35869
-----	CGTGTCAGGAGTGCATTTGC	-----	-----	-----	20	27827
-----	CGTGTCAGGAGTGCATTTG	-----	-----	-----	19	699
-----	-----	CTGAAACTATCTGAAGCATG	-----	-----	20	3168
-----	-----	TGAAACTATCTGAAGCATG	-----	-----	19	41
-----	-----	CTGAAACTATCTGAAGCAT	-----	-----	19	13
CTTTGAAGGTTTTGGGCTG	-----	-----	-----	-----	19	87
CCTTTGAAGGTTTTGGGCTG	-----	-----	-----	-----	20	60
TTTGAAGGTTTTGGGCTG	-----	-----	5' fixed	-----	18	15
-----	3' fixed	-----	-----	TCCAGAACGTACAATCTTCC	21	1
-----	-----	-----	-----	TCCAGAACGTACAATCTTCCGAA	25	1

(Berezikov et al. Genome Research, 2011.)

Single cell RNA-seq analysis



(Sandberg, Nature Methods 2014)

Long reads

Short reads



Long reads

Short reads

Long reads

