

Characterizing transcriptomes using ngs data

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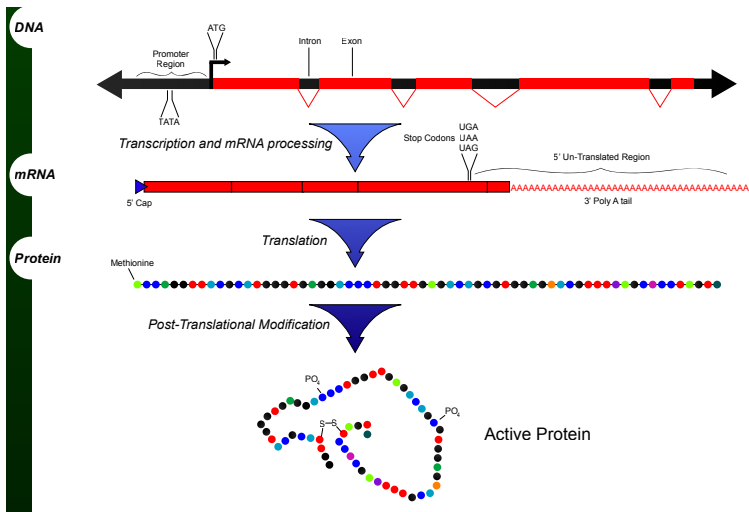
BILS/Scilife Lab/Uppsala University

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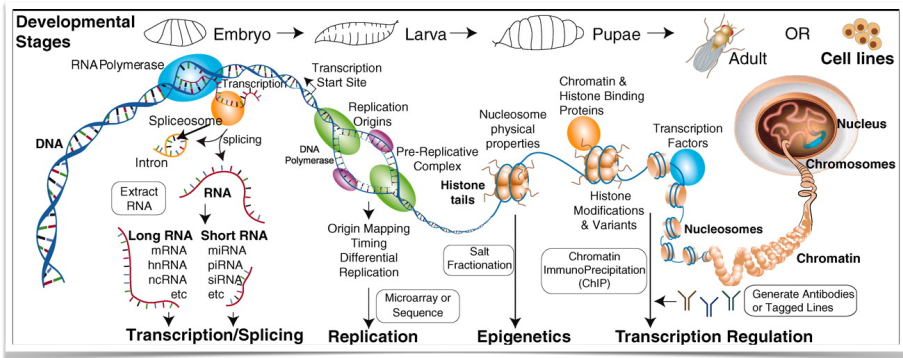
Outline

- 1 The transcriptome
- 2 RNA sequence technologies
- 3 RNA-seq analysis
 - Mapping based approach
 - Tools for working with ngs alignments
 - Gene expression from RNA-seq
 - de-novo assembly

The Central Dogma



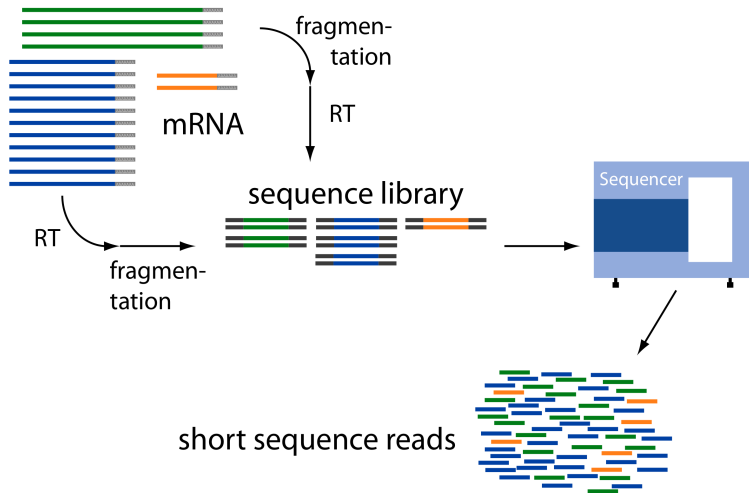
A more complex view



Transcriptomes vs genomes

- Dynamic, not the same over tissues and time points
- Smaller sequence space
- Less repetitive (but large gene families can be found)
- Fairly stable in size? (*eg.* 2-4 fold change among eukaryotes, whereas genome size can vary 1000-fold)
- Genes are often expressed in multiple different splice-variants
- RNA often from only one strand

NGS data



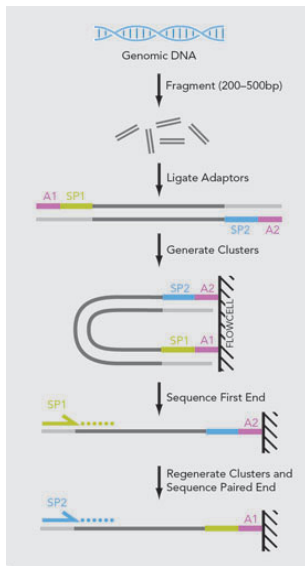
Machine output

```

@SRR038845.3 HWI-EAS038:6:1:0:1938 length=36
CAACGAGTTCACACCTTGGCCGACAGGCCCGGGTAA
+SRR038845.3 HWI-EAS038:6:1:0:1938 length=36
BA@7>B=>:>>7@7@>>9=BAA?;>52;>:9=8.=A
@SRR038845.41 HWI-EAS038:6:1:0:1474 length=36
CCAATGATTTTTTCCGTGTTTCAGAATACGGTTAA
+SRR038845.41 HWI-EAS038:6:1:0:1474 length=36
BCCBA@BB@BBBBB@B9B@=BABA@A:@693:@B=
@SRR038845.53 HWI-EAS038:6:1:1:360 length=36
GTTCAAAAAGAACTAAATTGTGTCAATAGAAAACCTC
+SRR038845.53 HWI-EAS038:6:1:1:360 length=36
BBCBBBBBB@@BAB?BBBBCBC>BBBAA8>BBBAA@

```


Pair-end (PE) sequencing



Pair-end reads

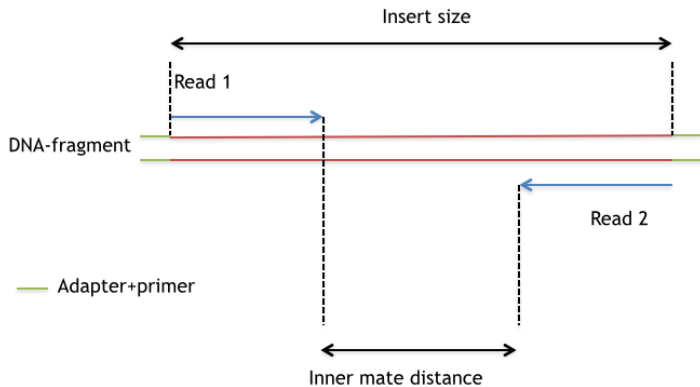
File format

- Two files are created
- The order in files identical and naming of reads are the same with the exception of the end
- The way of naming reads are changing over time so the read names depend on software version

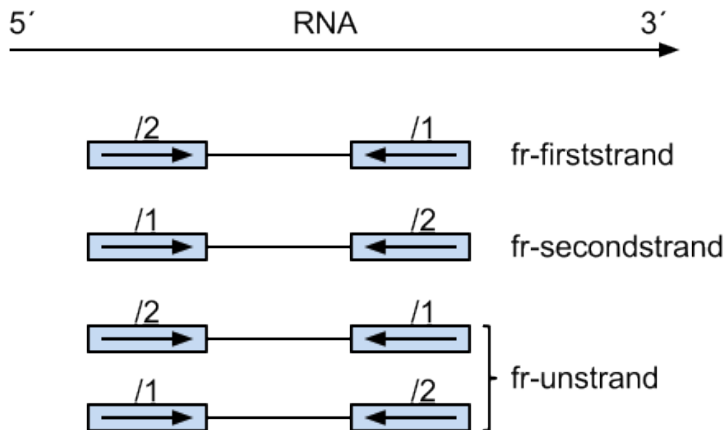
```
@61DFRAAXX100204:1:100:10494:3070/1
AAACAACAGGGCACATTGTCACTCTTGTATTTGAAAAACACTTCCGGCCAT
+
ACCCCCCCCCCCCCCCCCCCCCCCCCCCCCBC?CCCCCCCC@@CACCCCCA
```

```
@61DFRAAXX100204:1:100:10494:3070/2
ATCCAAGTTAAACAGAGGCCTGTGACAGACTCTTGGCCATCGTGTTGATA
+
_^_a^cccegcgghhgZc`ghhc^egggd^_[d]defcdfd^Z^OXWaq^ad
```

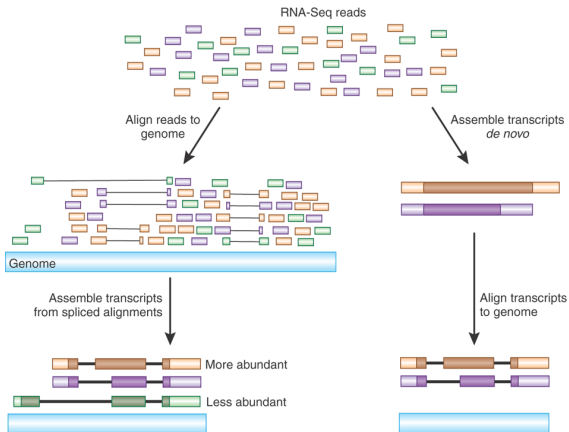
Pair-end data



Stranded or not



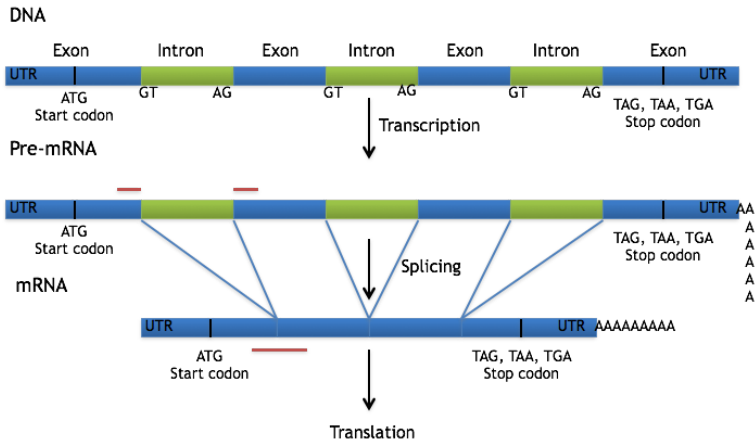
Two main routes for analysis



Haas & Zody (2010), Nature Biotechnology 28, 421–423

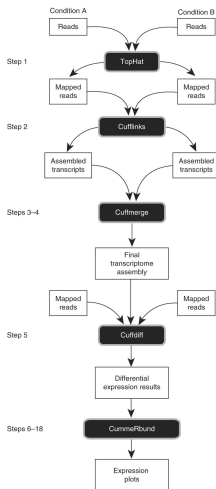
Aligning short reads from RNA to genomes

- Large number of programs available: Star, Tophat, Subread etc
- Important feature: Allow for spliced mapping



Example workflow

- Tophat: Aligns reads to genome (allows for spliced read mapping)
- Cufflinks: Extract transcripts from spliced read alignments
- Cuffmerge: Merge results from multiple Cufflinks results

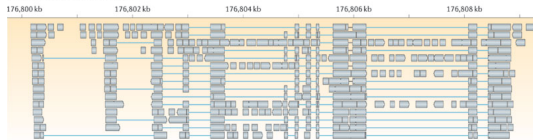
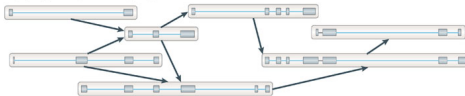
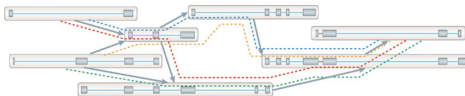


Trapnell *et al.* (2012), Nature Protocols 7, 562–578

Tophat

- ① Efficient and fast alignment to the genome using bowtie2
- ② Create a data base of putative splice junctions from the reads mapping in step 1
- ③ Map reads that did not map in step 1 run using the splice information

Cufflinks

a Splice-align reads to the genome**b Build a graph representing alternative splicing events****c Traverse the graph to assemble variants****d Assembled isoforms**

Samtools

- Program to work with ngs alignment files (SAM, BAM, CRAM)
- Can be used to view data, calculate basic info, extract subsets of alignments and convert between file formats
- <http://www.htslib.org>

Picard

- A set of Java command line tools with the same (or similar functionality as samtools)
- Note that even though they largely aim at doing similar functions Picard and Samtools is not always generating compatible file formats
- <http://broadinstitute.github.io/picard/>

IGV: Integrative Genomics Viewer

Viewer

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What's New

September 2014. The IGV iPad app can now be installed from the Apple App Store. *IGV for iPad* is a lightweight genomic data viewer that provides some of the functionality available in our regular desktop IGV. See the [IGV for iPad documentation](#) for details.

June 2014. We're hiring! See the [job description](#) on the Broad Institute careers website.

Overview

The Integrative Genomics Viewer (IGV) is a high-performance visualization tool for interactive exploration of large, integrated genomic datasets. It supports a wide variety of data types, including array-based and next-generation sequence data, and genomic annotations.

Downloads

Please [register](#) to download IGV. After registering, you can log in at any time using your email address. Permission to use IGV is granted under

Citing IGV

To cite your use of IGV in your publication:

Heiga Thorvaldsdóttir, James T. Robinson, Jill P. Mesirov. [Integrative Genomics Viewer \(IGV\): high-performance genomics data visualization and exploration.](#) *Briefings in Bioinformatics* 2012.

James T. Robinson, Heiga Thorvaldsdóttir, Wendy Winckler, Mitchell Gutman, Eric S. Lander, Gad Getz, Jill P. Mesirov. *Integrative Genomics Viewer.* *Nature Biotechnology* 29, 24–26 (2011)

Funding

Development of IGV is made possible by funding from the [National Cancer Institute](#), the [National Institute of General Medical Sciences](#) of the [National Institutes of Health](#), and the [Starr Cancer Consortium](#).

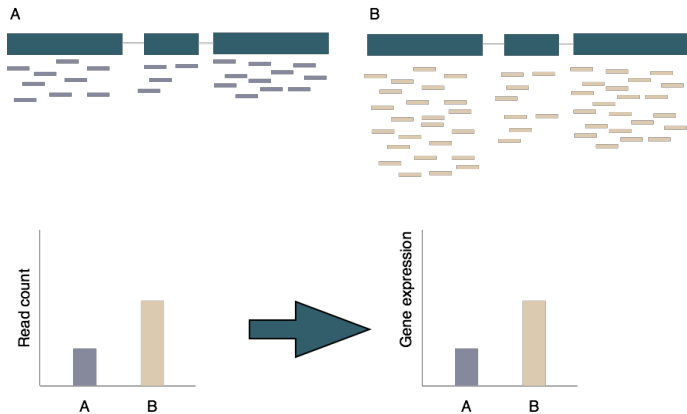
IGV participates in the [GenomeSpace Initiative](#), which is funded by the [National Human Genome Research Institute](#).



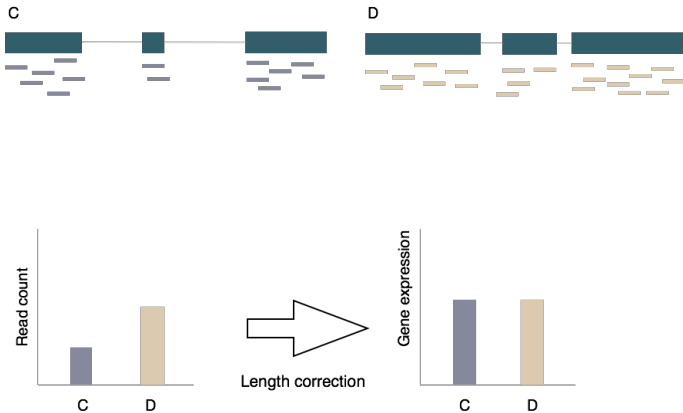
IGV: Integrative Genomics Viewer



From counts to gene expression



From counts to gene expression



Not all reads are the same

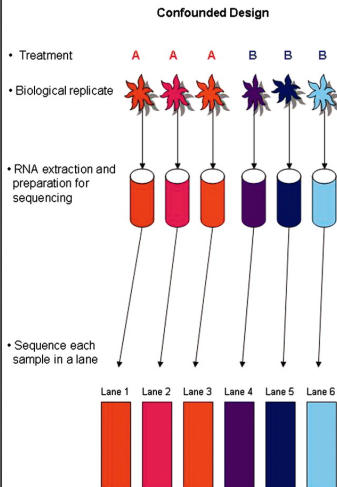
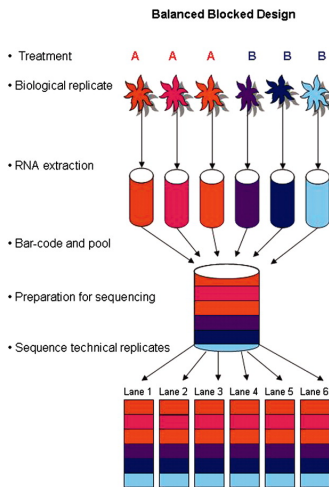
	union	intersection_strict	intersection_nonempty
	gene_A	gene_A	gene_A
	gene_A	no_feature	gene_A
	gene_A	no_feature	gene_A
	gene_A	gene_A	gene_A
	gene_A	gene_A	gene_A
	ambiguous	gene_A	gene_A
	ambiguous	ambiguous	ambiguous

from: <http://www-huber.embl.de/users/anders/HTSeq/doc/count.html>

Normalized expression Values

- Transcript-mapped read counts are normalized for both length of the transcript and total depth of sequencing.
- Count data is hence converted to: Reads/Fragments per kb of transcript length and million mapped reads (RPKM or FPKM)

Experimental design



Experimental design

- Count reads (convert to RPKM/FPKM?)
- Small number of reads (= low RPKM/FPKM values) often non-significant
- Remember that Fold change is not the same as significance

	Condition 1	Condition 2	Fold_Change	Significant?
Gene A	1	2	2-fold	No
Gene B	100	200	2-fold	Yes

Major challenges in relation to genome assembly

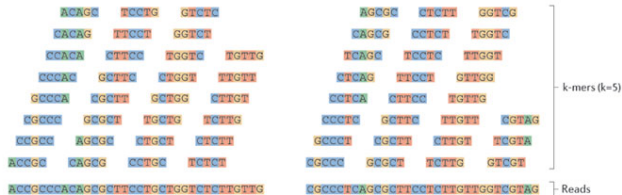
- Genes show different levels of gene expression, hence uneven coverage among genes
- Many genes are expressed in different isoforms
- As sequence depth increase detected number of loci increase. (What is actually expressed?)
- Sequence error from highly expressed genes might be seen more often than "true" sequences from lowly expressed genes

Several programs available

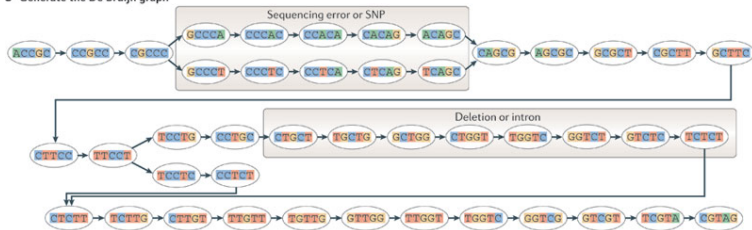
- SOAP-denovo TRANS
- Oases
- Trans-ABYSS
- Trinity

All of them uses de Bruijn graphs to cope with the data and many of them have been developed from a genome assembly program

Trinity

a Generate all substrings of length k from the reads

b Generate the De Bruijn graph



c Collapse the De Bruijn graph



Trinity

